Feasibility evaluation of amide proton transfer-weighted imaging in the parotid glands: a strategy to recognize artifacts and measure APT value

Yu Chen¹#, Xiaoqi Wang²#, Tong Su¹, Zhentan Xu¹, Yunting Wang³, Zhuhua Zhang¹, Huadan Xue¹, Zhizheng Zhuo², Yuanli Zhu¹, Zhengyu Jin¹*, Tao Zhang³*

¹Department of Radiology, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences, Beijing, China; ²Philips Healthcare, Beijing, China; ³Department of Stomatology, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences, Beijing, China

#These authors contributed equally to this work and are co-first authors.

*These authors contributed equally to this work.

Correspondence to: Zhengyu Jin. Department of Radiology, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences, No. 1 Shuai Fu Yuan, Dong Cheng District, Beijing 100730, China. Email: jinzy@pumch.cn; Tao Zhang. Department of Stomatology, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences, No. 1 Shuai Fu Yuan, Dong Cheng District, Beijing 100730, China. Email: drtzhang@126.com.

**Background:** The feasibility and image quality of three-dimensional (3D) amide proton transfer (APT)-weighted (APT w) in parotid tumor lesions have not been well established in previous studies. This study aimed to evaluate the utility of APT imaging in parotid lesions and glands.

**Methods:** Patients with parotid lesions received 3D turbo spin echo (TSE) APT w on a 3.0T scanner. Two radiologists, who were blinded to the clinical data, independently evaluated the APT w image quality using 4-point Likert scales (1= poor, 4= excellent) in terms of integrity and hyperintensity artifacts. An image quality selection protocol was built based on the two scores. Evaluable images (integrity score >1) and trustable images (integrity score >3 and hyperintensity artifacts score >2) were then enrolled for APT w value comparison between parotid lesions and glands.

**Results:** Forty consecutive patients were included in this study. Four patients were excluded due to severe motion (n=3) or dental (n=1) artifacts, and 36 patients received the APT sequence. Among these, more parotid tumor lesions (34/36, 94.4%) than normal parotid glands (23/31, 74.2%) revealed excellent integrity scores (score =4) (P=0.034). Most parotid tumor lesions (24/34, 70.6%) and glands (16/28, 57.1%) revealed no or little hyperintensity artifacts for diagnosis (scores 3 and 4). APT values of parotid lesions and glands in the evaluable groups were 2.11%±1.15% and 1.60%±1.56%, respectively, and the difference was not significant (P=0.197). APT values of parotid lesions and glands in the trustable groups were 1.99%±1.18% and 1.03%±1.09%, respectively, and the difference was statistically significant (P=0.018).

**Conclusions:** 3D APT w could be used to differentiate parotid tumors and normal parotid glands; however, the technology still needs to be improved to remove artifacts. In our study, most APT w images of tumor lesions in parotid glands had acceptable image quality, and these APT w images are feasible for diagnostic use.

**Keywords:** Parotid gland; magnetic resonance imaging (MRI); amide proton transfer (APT); image quality

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**Introduction**

Preoperative fine-needle aspiration biopsy results can provide important information on salivary gland mass lesions, such as whether the lesion is inflammatory or neoplastic and lymphoid or epithelial (1). However, sampling errors in heterogeneous tumors and non-diagnostic aspirate due to insufficient biopsy coverage occur in more than 10% of salivary gland tumors (2).

Modern imaging plays a key role in the differentiation of benign and malignant parotid lesions and selecting appropriate surgical strategies or radiotherapy (3). High-resolution ultrasonography, computed tomography (CT), and conventional magnetic resonance imaging (MRI) are the main imaging methods. MRI is superior in displaying the lesion interface and surrounding tissues without ionizing radiation. In the past, the roles of functional MRI, such as diffusion-weighted imaging (DWI) and dynamic contrast-enhanced (DCE) MRI, have been evaluated in parotid lesions (4-7). However, it is still challenging to differentiate benign and malignant lesions based on functional MRI. Therefore, advanced MRI techniques have been explored to provide biomarkers for the qualitative diagnosis of parotid lesions (8).

Amide proton transfer (APT) imaging is a non-contrast molecular imaging method. APT scopes the concentration of amide protons in tissue proteins and peptides by selectively saturating amide protons magnetization at 3.5 parts-per-million (ppm) and observing the affected water MR signal due to exchange between saturated amide protons and water protons. The much larger pool size of endogenous water protons (compared to the amide protons) and the continuous chemical exchange process has enabled the detection of free proteins and peptides with extremely low concentrations (9,10). APT MRI is a potential tool for tumor detection and characterization in the brain (11). More sophisticated clinical chemical exchange saturation transfer (CEST) applications exist that show a correlation between APT with therapy response (12-14) and patient outcome/survival (15).

Investigators in recent human studies have also reported preliminary APT findings in the breast (16-19), cervixes (20,21), prostate (22,23), and chest (8), demonstrating that it is an effective tool for tumor detection and characterization. APT values were higher in cancers than in normal surrounding tissues (22) and benign tumors (8). However, investigations applying the APT method in parotid tumors and normal parotid glands have not been well established in previous literature. There were a few reports about APT studies of parotid glands. Jing Yuan et al (11) demonstrated that parotid glands showed the highest APTw signal (~7.6% on average), while the APTw signals in other tissues were relatively moderate. Also, Yun Jung Bae et al. (24) demonstrated that the APTw signal was significantly higher in malignant parotid tumors than benign tumors (P<0.001). The APTw-signals exhibited an excellent area under the curve for predicting malignant tumors, which were markedly higher than the combined use of DWI and DWI-MRI (P=0.021 and 0.028, respectively). These studies used two-dimensional (2D) APT sequences, and only one slice was scanned for APTw. With a 2D readout, the shorter acquisition time is preferred in clinical settings, with only one single-slice acquisition. However, a three-dimensional (3D) readout could cover the whole lesion with multi-slice imaging and minimize relaxation loss between slices (25). Moreover, the commercially available 3D APT sequence for clinical use is easier for access and evaluation. Also, the 3D APT sequence is faster than the 3D version for five slices.

We expected APT MRI to provide head and neck image values to differentiate malignant tumors from benign lesions and normal tissues (11,26). Our first impression of APT application in parotid glands was unsatisfying because of relatively frequent image artifacts. However, a portion of parotid tumor APTw images showed correct diagnosis (24). Therefore, in this study, we investigated the feasibility of APTw imaging application in parotid glands and lesions in terms of image integrity and quality, and evaluated its ability to differentiate between lesions and normal tissues.

**Methods**

**Participants**

This study was conducted following the Declaration of Helsinki and was approved by an institutional review board. Written informed consent was obtained from the patients. Between December 2017 and October 2018, 40 consecutive patients with lesions in the parotid glands who planned to receive MR examinations were enrolled into this study. Four patients were excluded because the anatomic planning images were degraded by severe motion (n=3) or dental (n=1) artifacts. In total, 36 patients underwent 3D turbo-spin-echo (TSE) APTw MR imaging on a 3T MR scanner (Ingenia CX, Philips Healthcare, the Netherlands) equipped with dual radiofrequency (RF) transmit coils and a 32-channel head-coil for image acquisition. The patient selection flowchart is shown in Figure 1.
All patients underwent MR imaging with a clinical 3.0-Tesla MR system (Ingenia CX, Philips Healthcare, Best, the Netherlands) with a body coil for radiofrequency transmission and a 32-channel coil for signal reception.

Routine clinical imaging protocols were performed before the APT sequence, including axial T1 weighted turbo spin echo (T1w-TSE) [repetition time (TR), 701 ms; (echo time) TE, 14.772 ms; number of signal averaged (NSA), 2; slice thickness, 4 mm; matrix, 512×512], and axial T2 weighted turbo spin echo (T2w-TSE) (TR, 4,679 ms; TE, 114.912 ms; NSA, 2; slice thickness, 4 mm; matrix, 512×512). The anatomical images were also used to plan the field-of-view for the APT sequence with better localization.

The commercially available 3D APT sequence embedded the 3D TSE sequences. The 3D APT sequence parameters were as follows: slices, five; flip angle (FA) =90; TR/TE =2,000/8, the field of view, 250x250 mm²; slice thickness, 5 mm; voxel size, 2.50x2.53x5.00 mm³; TSE factor, 158; and Sensitivity Encoding (SENSE) factor, 1.6. The scan time of this sequence was 2 minutes and 40 seconds. Continuous RF saturation with no gap is essential for APT labeling. The MR scanner was equipped with two sets of transmitting RF amplifiers and coils, and the two transmission channels were turned on alternatively to produce four saturation pulses, each 500 ms in duration with an amplitude of 2 μT, to achieve a sufficient saturation of 2 seconds. The z-spectrum, with the saturation pre-pulse targeting at 3.5 ppm, 3.5±0.8 ppm, and −3.5 ppm, −3.5±0.8 ppm, and −1,560 ppm (according to the water proton frequency), was measured for each voxel. Images with saturation at the +3.5 ppm were acquired three times with an echo shift of 0.5 msec sequentially to calculate B0 field mapping (27,28) and then for z-spectrum correction (29).

APT value calculations were performed in the MR control console, according to previously published algorithms (29). B0 homogeneity was firstly measured and calculated via the echo shift in TSE acquisitions. With saturation at each offset, signals would be shifted to the corrected offsets in the z-spectrum based on the...
B0 field map. Signals at targeting offset $S(-3.5 \text{ppm})_{\text{corrected}}$ and $S(+3.5 \text{ppm})_{\text{corrected}}$ were calculated via the Lagrange interpolation, using the measured signals and the relative distances of measured offsets to targeting offsets.

APT weighting, measured as Magnetization Transfer Ratio (MTR) asymmetry, which is the signal difference between the corrected ±3.5 ppm images normalized to the −1,560 ppm image, was calculated according to the following equation:

$$APT \text{ weighting} = \frac{S(-3.5 \text{ppm})_{\text{corrected}} - S(+3.5 \text{ppm})_{\text{corrected}}}{S(-1560 \text{ppm})} \quad [1]$$

**Image analysis**

MR images and data were transferred to a workstation (Intellispace Portal; v. 10.1.0.64190; Philips Healthcare) for further analysis. One radiologist (YC) and one radiology scientist (XQW) with 10 years of head and neck experience independently reviewed all MR images. They were blinded to all patients’ clinical and histopathological data. Both the tumor lesions and parotid glands without lesions (from now on referred to as the parotid glands) were evaluated on APTw images, and both readers obtained consensus results.

The patients’ sex, age, and largest lesion diameter in axial images were recorded. The locations of lesions in the parotid gland were divided into upper, middle, and lower parts.

**Qualitative analysis**

Qualitative analysis was applied first to identify artifacts and evaluate image quality. We found strong artifacts at regions with large B0 inhomogeneity near air cavities and bone, where extrapolation fitting (rather than interpolation) had to be used for z-spectrum correction. Also, the low pass filter in APTw calculation spreads the extreme APTw values (>5% and <−5%) at the near skull and ear to parotid glands and lesions, which is our region of interest (ROI). In our study, we found that some APTw images of parotid glands and lesions were unsatisfactory for diagnosis due to these hyperintensity artifacts (APTw value >5%) or signal loss areas (APTw value <−5%). Hence, we need to recognize these image artifacts and decide if the samples could be enrolled in APTw value quantification.

The radiologists evaluated the parotid glands and lesions and drew the region of interest (ROI) on APTw mapping by manually delineating the whole tumor lesion or parotid gland concerning T2W images (Figures 2-4).

Firstly, a signal loss area of APTw images of lesions or parotid glands was evaluated within the coverage of ROI$_{w1}$ (Figures 2-4), in terms of the integrity scores based on a 4-point Likert scale: 4= excellent, the entire lesion or normal parotid gland was displayed; 3= good, more than 50% of the lesion or normal parotid gland was displayed; 2= moderate, less than 50% of the lesion or normal parotid gland was displayed; 1= poor, the lesion or normal parotid
gland was not displayed at all (Figure 5). The sample data was ruled out for further analysis if the integrity score was 0.

Secondly, image quality in the ROI of APTw images, in terms of the hyperintensity artifacts, was then evaluated based on a 4-point Likert scale: 4= excellent, tumor lesion or normal parotid gland could be recognized on APTw images without or with small hyperintensity artifacts; 3= good, hyperintensity artifacts impair less than 50% tumor lesion or normal parotid gland; 2= moderate, hyperintensity artifacts impair more than 50% tumor lesion or normal parotid gland; 1= poor, the entire tumor lesion or normal parotid gland is impaired by hyperintensity artifacts.

Samples of the parotid gland and lesions images with an intensity score of >1 and a hyperintensity artifacts score >1 were considered evaluable groups. After removing non-optimal image quality lesions or parotid glands (integrity score =2 and 3 and image quality scores =1 and 2), samples with intensity score >3 and hyperintensity artifact score >2 were considered evaluable groups.
Figure 5 An APTw image example of a poor integrity parotid lesion and gland. A 45-year-old male was found with a small round lesion at the left parotid gland and a normal parotid gland of the right side on T1WI (A). On APTw image (B), both the lesion and parotid glands showed no APTw signal. Therefore, the integrity of the lesion and parotid glands were considered as score 1 and the APTw value could not be measured.

were considered as trustable groups. The corresponding APTw values would be measured and recorded to compare both the evaluable and trustable groups in the next step.

Quantitative analysis

A second ROI was drawn on the APTw images (inside the ROIw), avoiding the visible hyperintensity artifacts and hypointensity area, and was recorded as ROIw2. The mean APTw values in ROIw2 of tumor lesions and parotid glands were all measured (Figures 2-4). The APTw values were then compared between tumor lesions and normal parotid glands in both the evaluable and trustable groups.

Statistical analysis

Statistical analyses were performed using SPSS software (IBM SPSS, version 19.0, SPSS Inc., Chicago, IL, USA). The correlation between the locations of lesions and integrity scores/hyperintensity artifacts were analyzed by Pearson correlation. Integrity and hyperintensity artifacts scores of tumor lesions and parotid glands were compared using Fisher’s exact test. The APT value between tumor lesions and parotid glands in both evaluable and trustable groups was compared using the t-test. All statistical tests were two-sided, and a P value less than 0.05 was considered statistically significant.

Results

Patients’ characteristics

Among the 36 patients who received the APTw examination, there were 18 men [58±12 years (range, 33–79 years)] and 18 women [45±15 years (range, 22–73 years)]. Data for five patients’ parotid glands without lesions were excluded because the structures were too small to evaluate. Hence, there were 36 lesions and 31 parotid glands included in the analysis.

Among the 36 lesions, five lesions located were in the upper part of the parotid gland, 23 lesions were located in the middle part of the parotid gland, and eight lesions were located in the lower part of the parotid gland. The lesions’ average diameter was 19.4±11.1 mm (range, 8.8–62.4 mm). Twenty patients received surgical treatments. The pathological results were as follows: 16 benign tumors, including 6 pleomorphic adenomas (PAs); 5 Warthin tumors (WTs), including 4 basal cell adenomas (BCAs) and 1 cystadenoma (CA); and four malignant tumors (MTs), including two mucoepidermoid carcinomas, one metastasis carcinoma, and one acinar cell carcinoma. The patients’ characteristics and selection flowchart is shown in Table 1 and Figure 1.

Qualitative analysis

In the integrity evaluation, 34 out of 36 lesions (94.4%) were considered for scoring as 4, and two lesions (5.6%, 2/36) were not shown completely and considered score 1. Twenty-three out of 31 normal parotid glands (74.2%) were considered for scoring 4, four (12.9%, 4/31) were score 3, one (3.2%, 1/31) was score 2, and three (9.7%, 3/31) were score 1. Parotid lesions had better APT image quality than parotid glands, and the difference between parotid tumors and parotid glands was significant (P=0.034).

After removing lesions or normal parotid glands with integrity scores of 1, 34 lesions and 28 parotid glands were
evaluated regarding the hyperintensity artifacts. 17.6% (6/34), 14.7% (5/34), 14.7% (5/34), and 55.9% (19/34) of parotid lesions were scored 1, 2, 3, and 4 respectively; and 21.4% (6/28), 21.4% (6/28), 7.1% (2/28), and 50% (14/28) of normal parotid glands were scored 1, 2, 3, and 4 respectively. There were no significant differences between the parotid lesions and normal parotid glands (P=0.729). The qualitative analysis results are shown in Table 2.

The intraclass correlation of integrity and hyperintensity artifacts scores of parotid lesions and glands by two independent readers were more than 0.9 (Table 3).

The patient selection flowchart is shown in Figure 1. Examples of parotid lesions and glands are shown in Figures 2-5. One patient (a 47-year-old male) received the B0 sequence. It was demonstrated that the region with stronger B0 variation, represented as the higher standard deviation in the B0 map, has more APT artifacts (Figure 6).

<table>
<thead>
<tr>
<th>Projects</th>
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<td>Patients received 3D APT sequence</td>
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<tr>
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<td>Male numbers</td>
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<tr>
<td>Female numbers</td>
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</tr>
<tr>
<td>Average diameter (mm)</td>
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<td>Severe motion</td>
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<td>Dental metallic artifact</td>
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<th>Projects</th>
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<th>Score 2 (%)</th>
<th>Score 3 (%)</th>
<th>Score 4 (%)</th>
<th>P value</th>
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<td>0</td>
<td>34 (94.4)</td>
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<td>Parotid glands (n=31)</td>
<td>3 (9.7)</td>
<td>1 (3.2)</td>
<td>4 (12.9)</td>
<td>23 (74.2)</td>
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<tr>
<td>Hyperintensity artifacts</td>
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<td></td>
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<td></td>
<td>0.729</td>
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<td>Parotid lesions (n=34)</td>
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<td>5 (14.7)</td>
<td>5 (14.7)</td>
<td>19 (55.9)</td>
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<tr>
<td>Parotid glands (n=28)</td>
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<td>6 (21.4)</td>
<td>2 (7.1)</td>
<td>14 (50.0)</td>
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</table>
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Quantitative analysis

After removing images with an integrity score of 1, the average APTw value of 28 samples of parotid lesions and 22 parotid glands in the evaluable groups were 2.11%±1.15% (range, 0.74–5.80%) and 1.60%±1.56% (range, −0.96% to 4.53%), respectively. There were no significant differences (P=0.197) in the direct comparison of APTw values between parotid lesions and glands.

Next, after removing non-optimal image quality in lesions or parotid glands (integrity score =2 and 3, and image quality scores =1 and 2), the average APTw value of 24 samples of parotid lesions and 14 parotid glands in the trustable groups were 1.99%±1.18% (range, 0.74–5.80%) and 1.03%±1.09% (range, −0.95% to 2.8%), respectively. There was a notable difference between lesions and parotid glands (P=0.018). The quantitative analysis results are shown in Table 4.

Discussion

This preliminary study investigated the first feasibility test of 3D TSE APTw MR imaging for parotid gland tumors. There are some advantages to the 3D TSE APTw MR method compared to previously-published 2D APT methods. In addition to the easy implementation with in-line post-processing, the slice with the biggest tumor diameter can also be retrieved. However, there are artifacts in APTw images based on the current technique. Only trustable samples, which can be identified via a measurable method, should be enrolled in preliminary studies. Hence, we developed a strategy in this study to initially evaluate
the image quality in terms of integrity and hyperintensity artifacts and then selected APT image samples with good image quality to continue the feasibility test.

Our result showed that the APT values of parotid gland lesions were higher than those of normal parotid glands. This is consistent with APT findings in other anatomies, including the brain, prostate, and cervical cancers. Previous research has demonstrated that higher APT values indicate a higher cellular proliferation rate, and our application in parotid glands is consistent with this assumption.

There are a few types of image artifacts. Firstly, the images have a low-resolution appearance after a low pass filter, and the hyperintensity artifacts from the low-fidelity APTw values near bone and air diffuse to parotid glands, thereby potentially triggering false positives. Higher-resolution APT imaging would be desirable with the less smoothing effect so that lesions and parotid glands would be less affected by surrounding hyperintensities. Second, large areas of liquefied necrosis, hemorrhage, or even large blood vessels often present high APTw values (30), which might be due to excessive water-exchangeable amide protons (i.e., proteins and peptides in the blood, cytoplasm, or other body fluids), could also be influenced by nuclear Overhauser Effect (NOE) in asymmetry analysis causing a high APT signal in fluids (31).

In our study, parts of pleomorphic adenoma with cystic area presented hyperintensity artifacts (Figure 7). Furthermore, the normal parotid gland’s intratumoral vessels always demonstrate hyperintensity on APTw images, as shown in the example in Figure 8. This finding

Table 4 Quantitative analysis of parotid lesions and normal parotid glands

<table>
<thead>
<tr>
<th>Projects</th>
<th>Parotid lesions</th>
<th>Parotid glands</th>
<th>P value</th>
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<td>Patient numbers</td>
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<td>22</td>
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<tr>
<td>APT value</td>
<td>2.11%±1.15% (0.74–5.80%)</td>
<td>1.60%±1.56% (−0.96% to 4.53%)</td>
<td>0.197</td>
</tr>
<tr>
<td>Trustable groups</td>
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<tr>
<td>Patient numbers</td>
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<td>14</td>
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</tr>
<tr>
<td>APT value</td>
<td>1.99%±1.18% (0.74–5.80%)</td>
<td>1.03%±1.09% (−0.95% to 2.8%)</td>
<td>0.018</td>
</tr>
</tbody>
</table>

Values given in parenthesis are the range of APT values.

Figure 7 An APTw image example of a hyperintensity artifact caused by cysts. A hyperintensity-signal lesion was found in the left parotid gland of 39-year-old female, as shown on T2WI (A). The pathological result was pleomorphic adenoma with cystic change. On the APTw (B) image, the integrity score was 4. The image quality in terms of hyperintensity artifacts scores was initially scored 4 by one radiologist and 2 by another. Eventually, the radiologists agreed on an image quality score of 2 because the hyperintensity in the cystic part of image was strongly affected by the artifacts diffused from the ear before the skull was hidden from the APT images.
is consistent with Chen et al.’s study of the liver (32), where the intrahepatic portal vein system and hepatic arteries were found to have high APTw values. In a recent study by Jiang et al. (33), tumor vessels in low-grade gliomas were shown always to exhibit high values on APTw images. Liquefaction necrosis, cystic degeneration, or large blood vessels could be identified in conventional anatomical MRI images (such as T2w and T1w); hence a reference to anatomical images is very useful to recognize APT image artifacts. These artifacts should be avoided when measuring APTw values for accurate interpretation (30). Thus, the image quality evaluation protocol is useful in screening for false positives and could serve as quality control.

The main challenge for APT imaging is B0 field inhomogeneity (20). Ideally, the APT calculation only needs to acquire images with saturation RF applied at ±3.5 ppm. However, the applied RF tuned at fixed frequencies could not saturate exactly at ±3.5 ppm due to B0 variation, while magnetization transfer ratio (MTR) asymmetry analysis depends on correct z-spectrum measurement. To improve B0 homogeneity, linear (and even second-order shimming) is used; however, B0 is still affected by bone, air, and other susceptible factors (11,24,34). Hence, a B0 map was measured (by acquiring three volumes with shifting echoes) to correct the z-spectrum with multiple offsets (3.5 ppm, 3.5±0.8 ppm, and −3.5 ppm, −3.5±0.8 ppm) acquired. To calculate the MTR asymmetry, the z-spectrum was shifted according to B0 at each voxel, and the measured points in the z-spectrum were used to interpolate the correct value. Our results showed the z-spectrum shift algorithm provided the necessary correction to generate APTw maps of parotid gland tumors (Figure 9). However, the APT value is complicated by B0, which is problematic in near air-tissue interfaces, resulting in strong artifacts in the near ear and oral cavities. Some tumor lesions and normal parotid glands had low image quality due to the poor B0 homogeneity (Figure 6). For B0 inhomogeneity that was too large to shift the z-spectrum within 0.8 ppm offsets, extrapolation fitting was used (rather than interpolation), and unacceptable errors would occur due to lack of accuracy during extrapolation fitting. We also found that the region with stronger B0 variation, represented as a higher standard deviation in the B0 map, had more APT hyperintensity artifacts, indicating that a large fluctuation in the B0 map is the key source of residual artifacts on APTw maps. RF labeling of amide proton saturation is significantly important for an APT sequence, and the variations in RF strength would lead to errors in APT calculation. Hence, the homogeneity of B1 plays an important role in defining correct saturations. There had been developments in the correction of B1 inhomogeneity (35), and integration of the new method might improve the diagnostic value of APT results.

Another source of artifacts in the APT image is the negligence of the NOE effect. The NOE effect affects the MTR asymmetry (similar to the APT effect), and there are errors in the APTw value if NOE is ignored in the calculation. Zhou et al. (36) showed that NOE positively
affects APTw in tumors, and thus, our report on APTw value measurement in tumors could be overestimated, and NOE has contributed to some high APTw values.

The APT sequence's saturation RF pulses in the present study lasted for 2 seconds, with an amplitude of 2 μT; and the resulting APTw value corresponds to these particular B1 settings (continuous labeling time and strength). The APTw value will change if the saturation RF pulses are changed.

A study by Takeda et al. showed that the parotid gland's saliva contains amino acids, pyruvic acid, and lactic acid (37). In normal tissues studied by Yuan et al. (11), the parotid glands showed the highest APTw signal, while the APTw signals in other tissues (submandibular gland, thyroid gland, and masseter muscle) were relatively moderate. The APTw value was influenced by the amide concentration and the proton exchange ratio between amide and water. Hence, the APTw value is affected by the acidity of tissue (11). Future investigations need to address amide concentration measurement more directly, which might have a greater clinical impact on parotid gland lesion diagnosis.

Our study had several limitations in the final data interpretation. Firstly, not all of the parotid gland tumors had pathological results. Secondly, image data with poor image quality were ruled out for quantitative analysis, which reduced the total sampling size. Thirdly, although the image integrity could be objectively evaluated, the current image quality analysis protocol regarding hyperintensity artifacts requires radiologists experienced in parotid anatomy and APTw image reading. Fourthly, significant differences between parotid lesions and healthy glands were only observed after removing data with non-optimal image quality. Fifthly, parotid glands’ contralateral sides without tumor lesions were considered the normal parotid gland groups. For these reasons, future prospective studies with a larger sample size are planned to characterize parotid gland tumors using the APT method. Lastly, we implemented the APTw protocol with a slice thickness of 5 mm to achieve a higher signal to noise ratio (SNR). However, the in-plane resolution and slice thickness restricted to use for smaller anatomy. Small lesions were not in the scope of this current preliminary study.

**Conclusions**

Our preliminary study of APT imaging for parotid tumors demonstrated that 3D APTw could be used to differentiate parotid tumors and normal parotid glands; however, the technology still needs to be improved to remove artifacts. Most APTw images of parotid glands’ tumor lesions had acceptable image quality, and these APTw images are feasible for diagnostic use.

**Acknowledgments**

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**Footnote**

**Conflicts of Interest:** All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/qims-20-675). The authors have no conflicts of interest to declare.

**Ethical Statement:** This study was conducted following the Declaration of Helsinki and was approved by an institutional review board. Written informed consent was obtained from the patients.

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