Introduction

Patellofemoral pain (PFP) is a common knee disorder in active young individuals comprising pain in and around the kneecap. Symptoms commonly occur during knee loading activities, such as running and stair climbing, and during sitting with the knees bent (1). Despite of a variety of treatment options, such as exercise therapy, patellar taping/bracing and foot orthoses, a large group of patients with persistent complaints remains (2-5). PFP has been
implicated as precursor of knee osteoarthritis (OA), but the exact pathophysiology remains unknown (6-8).

Pathophysiologic processes of the infrapatellar fat pad (IPFP), also known as ‘Hoffa’s fat pad’, have been proposed as a possible source of knee pain (9-13). The IPFP is a richly innervated, highly vascularized, intracapsular, extra-synovial structure in the anterior knee joint between the patella and femur, where it plays a biomechanical role (12,14-16). Structural changes of the IPFP, for example focal IPFP edema as sign of inflammation, have been pinpointed as precursor for structural knee OA, and a larger IPFP size was found in patients with patellofemoral OA (PFOA) (17-19). IPFP size could increase as a result of low-grade inflammation due to repetitive mechanical overload, as for example in Hoffa disease (20) or, hypothetically, can also be larger to begin with and predispose to pain without any pathophysiologic cause. In a PFP population no differences in the presence of focal edema of the IPFP between healthy control subjects and patients with PFP was demonstrated (21). IPFP size has been studied in OA (17,22), but has not been studied in PFP yet. Besides the biomechanical role, it is also suggested that the IPFP is an osteoarthritic joint tissue capable of modulating inflammatory responses in knee OA (11). This might also apply to PFP.

Dynamic contrast-enhanced magnetic resonance imaging (DCE-MRI) enables non-invasive evaluation of inflammation by measuring blood perfusion, which is known to increase in the presence of inflammation. DCE-MRI derived increased blood perfusion parameters are therefore considered imaging biomarkers of inflammation in various musculoskeletal tissues (23-28). To our knowledge, only one prior study applied semi-quantitative DCE-MRI in the IPFP of obese patients with knee OA and showed a correlation between knee pain and inflammation (28). In a previous study including the same study population as the current study, quantitative DCE-MRI analysis of the patella identified an increased patellar perfusion, contrary to the decreased patellar perfusion based on vascular alterations that was expected (29). Among the potential explanations of increased perfusion is the occurrence of an inflammatory process in which the IPFP plays an important modulating role (12).

Therefore, the aim of this study was to compare quantitative DCE-MRI blood perfusion parameters, as imaging biomarkers of inflammation, and volume of the IPFP between patients with PFP and healthy control subjects. A second aim was to explore if specific perfusion patterns exist in patients based on correlation of DCE-MRI perfusion parameters with clinical or MR imaging characteristics that potentially are related to inflammation or perfusion. Hypothetically, a larger IPFP volume and higher blood perfusion values are expected in patients with PFP than in healthy control subjects.

Methods

Study design and participants

In the current study, data was analyzed from a previously conducted cross sectional case-control study. Patients with minimum symptom duration of 2 months to a maximum of 2 years and healthy control subjects were included between January 2013 and September 2014. Patients who visited their general practitioner, physiotherapist or sports physician were included if diagnosed with PFP based on the presence of at least three of the following symptoms: pain while stair climbing, while squatting, while running, while cycling, sitting for a prolonged period with the knee flexed, or crepitus. Exclusion criteria were: previous PFP episodes more than 2 years ago, onset after trauma, defined pathological condition of the affected knee at present, or previous surgery or injury of the affected knee. Healthy controls were recruited from patients’ sports team members, friends, or colleagues. Exclusion criteria of controls were: history of PFP, surgery or injury of both knees, or first-degree relatedness with patients. Other exclusion criteria for both groups were: contra-indications for contrast-enhanced MRI and insufficient knowledge of the Dutch language. Patients and controls were aimed to match for age, gender, body mass index (BMI), and activity level. Full details of this study have been published elsewhere (21). This study was approved by our institutional review board, is conducted in accordance with the Helsinki Declaration and written informed consent was obtained from all participants. All patients and controls, aged 18–40 years, with DCE-MRI data available were included in the current analysis.

Image acquisition and measures

Participants underwent 3 Tesla MRI (Discovery MR750, GE Healthcare, USA) using a dedicated 8-channel knee coil (Invivo Inc., USA) at our institution. One knee, the (most) symptomatic knee of PFP patients was selected, or randomly chosen if both knees were equally painful or if both were asymptomatic (controls). The MRI protocol consisted of routine clinical proton density and T2-weighted fat-saturated sequences in three orthogonal planes,
and a sagittal 3D non-fat-saturated fast spoiled gradient-echo (non-FS FSPGR) sequence with a slice thickness of 0.5 mm.

DCE-MRI was acquired by a time-resolved imaging of contrast kinetics (TRICKS) sequence with anterior-posterior (AP) frequency encoding direction to avoid pulsation artifacts of the popliteal artery into the region of interest. MRI parameters were: in-plane pixel resolution 1.5 mm, slice thickness 5 mm, field of view 380×380×70 mm, acquisition matrix 256×128, 14 sagittal slices, 70% sampling in the phase direction, TE = 1.7 ms, TR = 9.3 ms, FA = 30°. The DCE-MRI protocol consisted of 35 phases of 10.30±0.07 s (constant within subject). Intravenous contrast administration of 0.2 mmol/kg gadopentetate dimeglumine (Magnevist, Bayer, Berlin, Germany), at a rate of 2 mL/s, was started after the first phase. Additionally, a non-fat-suppressed 3D FSPGR sequence with in-plane resolution of 0.3 mm × 0.3 mm and 0.5 mm slices was acquired before contrast administration for delineation of the patellar bone marrow.

In addition, participants completed a questionnaire on demographics, sports participation (yes/no) and knee complaints [Numerical Rating Score (NRS) pain score during rest and exercise, duration of symptoms, and function measured by the Anterior Knee Pain Scale (AKPS) 0–100 (30)]. Finally, a physical examination was performed in which the pressure pain threshold at the contralateral arm was tested as a measure of pain sensitization, according to a prior published method (31,32).

**Image analysis**

IPFP edema located centrally and superolaterally and joint effusion were already assessed as part of the Magnetic Resonance Imaging Osteoarthritis Knee Score (MOAKS) with several additional scoring items in a prior study (21,33).

The volume of interest (VOI) consisted of the whole IPFP with the following boundaries according to a recent study: the inferior patellar pole, femoral intercondylar notch, proximal patellar tendon, intermeniscal ligament, both menisci and the anterior tibia (15). VOIs were delineated in correspondence with the DCE-MRI data on the non-FS FSPGR sequence, which has previously been reported to be superior to fat-saturated images (Figure 1) (34). All VOIs were drawn by a senior radiology resident subspecializing in musculoskeletal imaging (RA van der Heijden) after careful consideration of the boundaries in the first 10 subjects together with a senior musculoskeletal radiologist (EHGO). DCE-MRI time points were registered using an automated rigid body registration with Elastix (35). Horos software (Horosproject.org, USA) was used to delineate the VOIs, register the non-FS FSPGR and DCE-maps, calculate the 3D volume and extract the perfusion parameters of the VOI with the DCE-tool (36).

Fitting a pharmacokinetic model to the data enables extraction of quantitative DCE-MRI parameters which (to a greater or lesser extent) reflect physiological phenomena such as blood flow, blood volume, and extravascular permeability (37). Tofts’ pharmacokinetic model has shown to be the most accurate model for patellar bone (38). Tofts’ extended model is more suitable for highly vascularized structures, such as the IPFP, by adding the vascular term $V_p$ (37). The arterial input function (AIF) was estimated in each participant using a ROI in the popliteal artery. All fitted AIF's were visually checked. Quantitative DCE-MRI perfusion parameters $K_{trans}$, $K_{ep}$, $V_e$ and $V_p$ were extracted by fitting the Tofts’ extended pharmacokinetic model (Figure 2) (39). $K_{trans}$ reflects the volume transfer constant into the tissue compartment, $K_{ep}$ describes the rate constant back into the vascular component, $V_e$ the extravascular extracellular space and $V_p$ the vascular fraction.

**Statistical analysis**

The Shapiro-Wilk test was used to evaluate normality of the distribution of the parameters. Independent sample t-tests and chi-square tests, or Mann-Whitney U tests if data distribution was not normal, were applied to investigate
differences in baseline characteristics between groups.

The DCE image analysis resulted in a mean value for the perfusion parameters within each VOI. The mean and standard deviation over all subjects were calculated for control subjects and PFP patients separately. Differences in variance of volume and perfusion parameters across subjects were tested with Levene’s test. Volume followed a normal distribution. All perfusion parameters showed a normal distribution of residuals after logarithmic transformation and, accordingly, regression analyses could be performed. Differences in volume and perfusion parameters between groups were compared by linear regression analyses, adjusted for age, gender, BMI, and sports participation. Furthermore, possible patient subgroups were explored by multivariate linear regression analysis of the following variables: pain during rest and during exercise, duration of complaints, presence of sitting pain, function, pain pressure threshold contralateral arm as sensitization measure, presence of IPFP edema centrally and superolaterally and joint effusion. P values <0.05 were considered to be statistically significant for the main linear regression analyses. Results are presented as mean differences with 95% confidence intervals. For the subgroup analyses, a lower P value of <0.01 was applied due to multiple testing. All analyses were performed with SPSS v25 (IBM, USA).

Results

Population and patient characteristics

In a prior study, 64 patients with PFP and 70 control subjects aged 14–40 years were included (Figure 3). DCE-MRI was only acquired in adults and image quality was sufficient in 35 adults PFP patients and 44 adult control subjects. Mean age was 26.1 (range, 18–40, SD 5.0) years, mean BMI was 24.1 (SD, 3.4) kg/m² and 49% (39) was female. The BMI was significantly higher in the patient group (Table 1). Patients reported a mean duration of complaints of 11.2 months and 45.7% (16) reported bilateral pain. Centrally located moderate to severe IPFP edema was present in one patient and two control subjects. Superolateral IPFP edema was present in 16 patients and 19 control subjects. Medium to large effusion (corresponding with MOAKS grade 2–3) was present in four patients and seven control subjects. There were no significant differences in the presence of these features.

Volume and DCE-MRI parameters

Due to a lack of a plateau phase in the time intensity curve, the fitting algorithm might not provide valid values of \( V_e \) and subsequently \( K_{ep} \) and therefore those parameters were not shown.

Mean IPFP volume was 26.04 (4.18) mL in control subjects and 27.52 (5.37) mL in patients. Median \( K_{trans} \) was 0.017 (0.016) min⁻¹ in control subjects and 0.016 (0.020) min⁻¹ in patients. Median \( V_p \) was remarkably higher in patients than control subjects, respectively 0.00037 (0.00039) and 0.00029 (0.00033), but this difference was not statistically significant (P=0.10). Mean volume and perfusion parameters did not differ between groups (Table 2, Figure 3).

\( V_p \) was the only perfusion parameter for which Levene’s test showed a significant difference in variance across groups (P=0.035). Further visual inspection of the boxplots showed two outliers in the patient group, which most likely account for the difference in variance (Figure S1: boxplot without the largest patient outlier).

The patient subgroup analyses showed that presence of joint effusion was significantly associated with elevated \( K_{trans} \) (P=0.002) and \( V_p \) (P=0.004), but not with volume (P=0.41). Remarkably, this association was not present in control subjects. None of the other subgroup analyses...
revealed significant associations for respectively volume/$K_{\text{trans}}/V_p$: pain during rest $P=0.76/0.74/0.31$; during exercise $P=0.07/0.25/0.51$; duration of complaints $P=0.78/0.30/0.14$; presence of sitting pain $P=0.29/0.23/0.24$; function $P=0.57/0.88/0.48$; pain pressure threshold contralateral arm as sensitization measure $P=0.82/0.54/0.27$; presence of IPFP edema centrally $P=0.11/0.44/0.56$; presence of IPFP edema superolaterally $P=0.82/0.75/0.54$.

**Discussion**

In this study, volume and quantitative DCE-MRI blood perfusion parameters $K_{\text{trans}}$ and $V_p$ of the IPFP were compared between healthy control subjects and patients with PFP. In contrast to our hypothesis, no significant differences were found in volume or DCE-MR blood perfusion parameters, as imaging biomarkers of inflammation, of the IPFP between healthy control subjects and patients with PFP. Furthermore, patient subgroups were explored in search of an association of DCE-MRI parameters with clinical characteristics and MR features potentially related to volume or perfusion/inflammation. Only effusion was significantly associated with higher $K_{\text{trans}}$ and $V_p$ in patients with PFP.

In recent studies, inconsistent results were found in OA populations regarding IPFP volume. One study found an association between a larger volume and fewer structural abnormalities in patients with clinical knee OA, suggesting a protective role of a larger IPFP (40). Two other studies did not find a correlation between IPFP volume and
symptomatic or radiographic knee OA (17,41). A fourth study, specifically focusing on PFOA, demonstrated that individuals with PFOA had a larger IPFP than controls, and a larger IPFP volume was directly related to pain (42). Since age presumably influences fat pad volume in patient with OA, the current volumes cannot be directly compared with the prior study including an older population of patients with PF OA (43).

The single prior study that applied DCE-MRI, in a semi-quantitative manner, to investigate IPFP inflammation found a relation between their perfusion derived inflammation marker and knee pain in obese patients with knee OA (28). DCE-MRI had not yet been applied in the IPFP of patients with PFP and healthy control subjects.

The lack of a difference between patients with PFP and control subjects might be explained by a still preserved tissue homeostasis in PFP without induced inflammatory response. In order to determine if a certain patient subgroup with explicit inflammation could be identified, additional exploratory subgroup analyses were conducted focusing on clinical characteristics and MR features potentially related inflammation. For instance, a larger size and higher

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Patients (n=35)</th>
<th>Controls (n=44)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Female gender, n (%)</td>
<td>18 (51.4)</td>
<td>21 (47.7)</td>
<td>0.74</td>
</tr>
<tr>
<td>Age (years), mean (SD)</td>
<td>26.4 (5.6)</td>
<td>25.9 (4.6)</td>
<td>0.53</td>
</tr>
<tr>
<td>BMI (kg/m²), mean (SD)</td>
<td>25.1 (3.8)</td>
<td>23.3 (2.8)</td>
<td>0.01</td>
</tr>
<tr>
<td>Sports participants, n (%)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>During inclusion</td>
<td>24 (68.6)</td>
<td>34 (77.3)</td>
<td>0.39</td>
</tr>
<tr>
<td>Before onset of pain</td>
<td>32 (91.4)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Pain (NRS), mean (SD)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>During rest</td>
<td>3.9 (2.7)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>During strain</td>
<td>6.3 (2.4)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Duration of complaints, mean (SD)</td>
<td>11.2 (6.3)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Bilateral pain, n (%)</td>
<td>16 (45.7)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Sitting pain, n (%)</td>
<td>27 (77.1)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>AKP function score, mean (SD)</td>
<td>68.6 (11.0)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Pain pressure threshold arm, mean (SD)</td>
<td>50.3 (13.8)</td>
<td>56.1 (14.3)</td>
<td>0.07</td>
</tr>
<tr>
<td>IPFP edema central (moderate to severe), n (%)</td>
<td>1 (2.9)</td>
<td>2 (4.5)</td>
<td>0.59</td>
</tr>
<tr>
<td>IPFP edema superolateral, n (%)</td>
<td>16 (45.7)</td>
<td>19 (43.2)</td>
<td>0.50</td>
</tr>
<tr>
<td>Effusion (medium to large), n (%)</td>
<td>4 (11.4)</td>
<td>7 (15.9)</td>
<td>0.50</td>
</tr>
</tbody>
</table>

BMI, body mass index; NRS, Numerical Rating Score; AKP, Anterior Knee Pain; IPFP, infrapatellar fat pad.

<table>
<thead>
<tr>
<th>Measures</th>
<th>Patients (n=35)</th>
<th>Controls (n=44)</th>
<th>Mean difference (95% CI)</th>
<th>Adjusted P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Volume (mL)</td>
<td>27.52 (5.37)</td>
<td>26.04 (4.18)</td>
<td>1.48 (–0.66, 3.62)</td>
<td>0.07</td>
</tr>
<tr>
<td>K\text{\textit{trans}} (min\textsuperscript{-1})</td>
<td>0.016 (0.020)</td>
<td>0.017 (0.016)</td>
<td>–1.12 (–13.47, 11.21)</td>
<td>0.98</td>
</tr>
<tr>
<td>V\text{\textit{p}}</td>
<td>0.00037 (0.00039)</td>
<td>0.00029 (0.00033)</td>
<td>1.10 (–0.65, 2.85)</td>
<td>0.10</td>
</tr>
</tbody>
</table>

VOI, volume of interest.
perfusion of the IPFP could be expected in patients with other proposed signs of inflammation, such as Hoffa edema or effusion. Furthermore, we hypothesized that low-grade inflammation resulting in higher perfusion and/or increased volume would be associated with more pain and worse function or would have a more systemic effect resulting in longer symptom duration or presence of sensitization. Finally, an increase in size or vascular changes could also have been the answer to the enigma of patients with PFP, which explicitly exhibit pain during sitting with the knees bent (29,44,45). In the end, none of these variables were significantly associated with volume or perfusion parameters besides effusion. Effusion was significantly associated with $K_{trans}$ and $V_p$, indicating a larger vascular fraction and more blood inflow in patients. Further evaluation revealed that these associations were not present in control subjects.

This implies that joint effusion is only associated with inflammation and neovascularization in patients with PFP. These results are only speculative, though, due to the low number of subjects per group. A potential explanation for the difference between groups might lie in a different mechanism of effusion, implying an inflammatory pathway in patients only. In control subjects one possible explanation might be a mechanical pathway.

In this study, for the first time, the IPFP volume and IPFP blood perfusion were quantitatively analyzed by DCE-MRI in order to unravel the role of the IPFP in the pathophysiology of PFP. A strength of this study is the inclusion of healthy control subjects next to patients with PFP. Furthermore, the current study applied quantitative assessment of DCE-MRI perfusion values, which offers more robust parameters that directly represent the microvasculature physiology, in contrast to semi-quantitative analysis.

A potential limitation might be the lack of B1+ or pre-contrast T1 map, which led to the use of a literature based fixed T1(0) value of 1,443 ms. We do not expect this to have affected the outcome, since no differences in native T1 variability were expected between groups. Furthermore, a dedicated transmit/receive knee coil with relatively homogeneous B1 field was used. Second, quality of DCE-MRI imaging data was not sufficient in nine patients and six control subjects due to artifacts at the beginning of the study. We do not expect this to have influenced our conclusions, as the baseline characteristics of these participants did not differ from the participants in which the DCE-MRI was sufficient. Due to time constraints in our MR protocol, DCE-MRI acquisition did not last long enough for the time intensity curve to reach plateau phase. As this might potentially lead to unreliable estimates of $V_c$ and subsequently also of $K_{trans}$, only the robust parameters $K_{trans}$ and $V_p$ were presented. This is sufficient for our research purpose as these two are the most important parameters to identify increased perfusion.

ROI delineation was done by a single observer only, which leaves the variability introduced by having an alternative observer unknown. A previous study stated that the inter-observer variability is low when a large ROI is used (46). This was done in quantitative MRI of cartilage, but given our experience in both cartilage and other structures like the fat pad we found these comparable. Furthermore, the boundaries of the ROI were discussed in depth with a senior musculoskeletal radiologist (EHGO).

Since a group-wise AIF is not available within the DCE-tool, we used a subject-specific AIF, despite the probability of not capturing the arterial bolus given the low temporal resolution of 10 seconds. Therefore, all AIF curves were visually checked and appeared to capture the bolus peak adequately. Another point to notice is that small, yet significant differences could have been left undetected due to the large inter-subject variability. Power analysis was not feasible in advance, due to the lack of knowledge regarding effect sizes. In future research including a larger number of subjects would be advised in order to be able to draw firm conclusions. With regard to our results, we do not think this would affect the main outcome as inter-subject variability is equally present in both groups and the variance was not statistically significantly different between groups, except for $V_p$. The clinical relevance of these potentially missed small differences would also be questionable given the small effect sizes and the fact that patients had a lower mean of $K_{trans}$ and $V_p$, which does not concur with the suspected increased blood perfusion accompanying inflammation. Finally, some of the subgroup analyses were theoretically underpowered due to the low prevalence of features. If these would have been the key features for the incidence of PFP, though, a higher prevalence would have been expected to start with. In general, due to the cross-sectional design no causal relations could be studied, but only associations. Future research might try to achieve a complete picture of the perfusion in the knee joint. For now, this is quite challenging, because different tissues require different pharmacokinetic models.

In conclusion, patients with PFP and healthy control subjects do not demonstrate a significantly different volume or blood perfusion of the IPFP. Thus, it seems that higher
IPFP blood perfusion measured by DCE-MRI, as imaging biomarker of inflammation, and larger volume are not associated with PFP.

Acknowledgments

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/qims-20-441). EHGO serves as an unpaid Associate Editor of Quantitative Imaging in Medicine and Surgery. Dr. RA van der Heijden reports grants from European Society Skeletal Radiology, during the conduct of the study; Dr. M van Middelkoop reports grants from Erasmus University Rotterdam, during the conduct of the study; Dr. SMABZ reports grants from Dutch Arthritis Foundation, during the conduct of the study; Dr. EHGO reports grants from RSNA, during the conduct of the study. The other authors have no conflicts of interest to declare.

Ethical Statement: The study was approved by the institutional ethics committee of the Erasmus University Medical Center (No. 2012-342). Written informed consent was obtained from the patient for publication of this study and any accompanying images.

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**Figure S1** Boxplot of $V_p$ multiplied by 1,000 with exclusion of the biggest patient outlier. The * means the remaining outliers.