Introduction

Chronic kidney disease (CKD) is a global health issue, since 11–13% of the world population can be diagnosed with CKD in different severity stages (1). The syndrome is characterized by several nutritional and metabolic derangements, that become clinically evident in most advanced stages. A number of metabolic conditions commonly observed in severe CKD (e.g., acidosis,
inflammation, oxidative stress, and impaired insulin/insulin-like growth factor axis function etc.), chronic comorbidities, and renal replacement therapy (RRT) (e.g., dialysis) per se have a negative impact on nutritional status, as they can induce muscle wasting through the alteration of the balance between muscle protein anabolism and catabolism (2).

The evaluation of nutritional status is usually based on a comprehensive assessment including patient history, physical examination, evaluation of nutrient intake, biochemical markers and screening methodologies that provide a global picture of patients' protein and energetic reserve, as well as their nutritional risk (3). An important part of this evaluation is the assessment of body composition, which refers typically to the quantification of adipose tissue and, more importantly, of muscle mass, aiming for an early diagnosis and monitoring in patients at risk of muscle loss.

Nowadays, several methodologies are available for the estimation/measurement of muscle mass (Table 1). Some of these techniques, such as anthropometry, bioimpedance analysis (BIA) and, with some limitations, dual energy X-ray absorptiometry (DXA) are available for routine patient assessment (4). Other methodologies—mainly imaging techniques, such as computed tomography (CT) and magnetic resonance imaging (MRI)—are currently used only in research studies and represent the gold standard (5). More recently, muscle ultrasound (US) has been studied as an alternative tool for the assessment of muscle mass at the bedside in different clinical settings, including CKD and acute kidney injury (AKI) (6-14). While anthropometry and BIA are surrogates of muscle mass, imaging techniques should be preferred for a precise quantification of both adipose and muscle tissues.

In this paper, we provide an overview of the different methods currently available for the assessment of body composition in renal disease, focusing on imaging techniques and their applicability for the diagnosis of muscle wasting.

**Impact of muscle wasting in renal disease**

Skeletal muscle is vital to mobility, posture, strength and balance as it allows the performance of physical activities of daily living (15). In addition, it is also a pivotal metabolic and homeostatic organ, via crosstalk with other organs systems (16). Most importantly, it plays a key role in protein metabolism as a source of amino acids when protein intake is insufficient, thus preserving the protein content of other essential organs (16,17). However, when muscles undergo chronic catabolism to supply amino acids for other metabolic purposes, a reduction in lean body mass (LBM) is observed. This has potentially serious clinical consequences such as muscle weakness, impaired physical function, increased morbidity and mortality (18). In fact, as low as 10% loss of LBM is associated with increased risk of death, due to impaired immune response and increased susceptibility to infections, while a 40% reduction is incompatible with life (19).

Decreased muscle mass is a frequent finding in patients with end-stage kidney disease (ESKD) on hemodialysis (7), with muscle loss occurring earlier and more severely in this population in comparison to age-matched controls (20,21). This phenomenon has been associated to nutritional problems, chronic diseases, a sedentary lifestyle and drug-related side effects (22,23), and its prevalence seems to be strongly related to the worsening of kidney function (24). The consequences of muscle loss are not only physically debilitating. In fact, many studies in the past decades have also linked muscle wasting in ESKD patients on dialysis with worse quality of life, depression, protein energy malnutrition, cardiovascular complications, as well as...
with increased hospitalization and mortality (25-29). In addition, the negative consequences of muscle wasting are not only a concern to patients on dialysis, as also in kidney transplant recipients sarcopenia, which is characterized by a loss of muscle mass and function, correlates with increased mortality, graft failure, and more frequent postoperative complications (30-33).

There is strong evidence linking muscle wasting to worse prognosis in CKD and other non-communicable diseases states, such as chronic obstructive pulmonary disease (COPD), or chronic heart failure (CHF) (34-38). Despite the diverse pathophysiology of these diseases, all are associated with increased protein catabolism through the upregulation of the ubiquitin-proteasome system (UPS) (35,39-42).

Most of these pathological conditions are associated with various degrees of chronic inflammation, which plays a critical role in the onset of muscle atrophy and malnutrition. During the physiologic process of atrophy, the degradation of myofibrillar and soluble proteins is accelerated. However, in systemic wasting conditions, protein synthesis also decreases, leading to a negative balance between degradation and synthesis, which will eventually result in muscle loss, reduced body weight, weakness and disability (43).

Physical disability is characterized by difficulties in performing activities of daily living, worsens quality of life and increases the risk of death (44).

**Pathophysiology of muscle wasting in CKD**

The pathogenesis of muscle wasting in CKD is multifactorial and is characterized by a persistent vicious cycle between muscle wasting and its complications (Figure 1). Based on currently available evidence, the causes of muscle wasting in CKD can be classified into two groups: (I) causes related to kidney disease *per se*; and (II) causes related to chronic low-grade inflammation, typical of patients on dialysis but also present in earlier stages of CKD (45,46). Factors related to kidney disease that contribute to the loss of skeletal muscle mass (SMM) include inadequate spontaneous nutrient intake, metabolic acidosis, vitamin D deficiency, insulin resistance, low physical activity, proteinuria and hyperparathyroidism (2).

Inadequate nutrient intake is quite frequent in these patients, and is the most important factor that contributes to the development of muscle wasting. Progressive loss of appetite (anorexia) begins already in early stages of...
CKD (47), and worsens in parallel to the loss of kidney function. In this clinical setting, anorexia is a frequent consequence of taste abnormalities, accumulation of uremic toxins, dysregulation of gastrointestinal homeostatic mechanisms, altered blood concentration of appetite regulators and deranged hypothalamic output (47). In addition, CKD patients on unsupervised dietary nutritional treatment (the so called low-protein diets) usually undergo prolonged restrictions of protein intake, as well as of phosphorus, potassium and sodium, that can be deleterious if accompanied by inadequate energy intake (48,49). Furthermore, frailty, poverty, advanced age and multiple acute or chronic comorbidities may also contribute to suboptimal nutrient intake in ESKD (30,50-52).

Metabolic acidosis is a very frequent complication of ESKD, is associated with increased mortality (53), and plays an important role in the stimulation of protein catabolism (54). In fact, metabolic acidosis increases intracellular protein degradation by activating both the caspase-3 and the UPSs, promotes the oxidation of branched chain amino acids, reduces protein synthesis and favors insulin and GH resistance, thus leading to negative protein balance (55,56). These effects can be reversed by bicarbonate administration (57,58).

Insulin resistance is one of the most important metabolic challenges in diabetic patients with CKD, who suffer from more severe protein degradation and loss of LBM in comparison to non-diabetic patients (59,60). However, insulin resistance can be observed also in non-diabetic patients on dialysis, and is associated with increased protein catabolism mediated primarily by the ubiquitin-proteosome pathway, with insulin resistance acting through a decrease in muscle phosphatidylinositol 3 kinase (PI3K) (61). In addition to metabolic acidosis, vitamin D deficiency also contributes to the development of insulin resistance by affecting pancreatic insulin secretion (62,63). Moreover, vitamin D deficiency can directly affect muscle synthesis (64).

Among other causes for the development of muscle wasting, chronic inflammation is a frequent finding in ESKD patients. It may contribute to both an increase in nutritional needs and to anorexia, through an imbalance between the orexigenic/anorexigenic mechanisms that control the energetic homeostasis of renal patients (51,52). Several factors are thought to contribute to the pathogenesis of inflammation in ESKD: worsening of renal function, which reduces the elimination of pro-inflammatory cytokines and uremic toxins; the presence of comorbidities; and factors related to the dialytic treatment per se, such as membrane and dialysis fluid bioincompatibility (65).

Furthermore, the available evidence suggests a pivotal role of the gastrointestinal tract in the pathogenesis of chronic inflammation in CKD, as a consequence of intestinal dysbiosis and barrier disruption (66-68). In healthy subjects, beneficial bacterial species characterized by a predominant saccharolytic metabolism outweigh pathobionts. They are responsible for fermenting complex carbohydrates and producing short chain fatty acids, such as acetate, butyrate and propionate, that serve as fuels for the intestinal epithelium and have also favorable immunomodulatory effects (68,69). The uremic milieu and the reduced availability of non-digested complex carbohydrates, with the latter being a common consequence of the reduced intake of fibers characteristic of CKD/ESKD patients, are responsible for negative effects both on the resident microbial population (i.e., dysbiosis,) and on the structure/function of the gastrointestinal tract, enhancing the permeability of the intestinal barrier (66,68,70). This dysbiotic environment is characterized by a switch towards a more proteolytic metabolic profile, leading to an increase in protein fermentation (putrefaction) and the ensuing generation of increased amounts of potentially toxic compounds (i.e., ammonium, thiols, phenols, indoles) that are absorbed into the bloodstream (68,70). In addition, increased urea influx from the blood into the gut promotes urea hydrolysis by microbial species that contain urease to form large quantities of ammonia, which is converted to ammonium hydroxide (71). Ammonia and ammonium hydroxide are toxic compounds known to disrupt the integrity of the gut epithelial barrier, also increasing its permeability (68). The deranged and more permeable intestinal barrier may facilitate bacterial translocation, i.e., the passage of bacteria or their structural component lipopolysaccharide (LPS) from the gut lumen to the blood (68,70), triggering the inflammatory cascade. Tumor necrosis factor alpha (TNF-α), interleukin 6 (IL-6), IL-8, and interferon gamma (IFN-γ) are in fact mentioned among the most frequently observed indicators and stimulators of muscle proteolysis, while C-reactive protein (CRP) seems to be a useful and inexpensive, although nonspecific, marker of systemic inflammation (25,72).

Finally, physical activity is markedly reduced in hemodialysis patients in comparison to age-matched controls (73,74). Reasons for reduced physical activity are fatigue on dialysis days, lack of time and motivation, physical problems and pain (75,76). In this clinical setting, physical inactivity represents a modifiable risk factor for the development of muscle loss and, in addition to
the promotion of muscle atrophy, it may cause a further increase in the already high level of cardiovascular risk in these patients (74,77). Physical exercise, even in dialyzed patients, ameliorates depression and improves quality of life, appetite and energy supply (77).

**Methods to assess body composition**

**Anthropometry**

Anthropometry can be considered the first step in the evaluation of the nutritional status of CKD patients. It includes the measurement of body dimensions, i.e., body weight, height, skinfold thickness and circumferences of limbs, waist and hips. From these measures, it is possible to calculate further parameters such as the body mass index (BMI) (weight/height², kg/m²), waist/hip ratio, and surface area.

Due to low reproducibility and reliability on an individual basis, anthropometric measurements are sometimes considered to be less important than other measurements requiring more sophisticated techniques (78). However, they are simple to perform, noninvasive, cheap, provide immediate results and are applicable on a large scale. Thus, the use of specific tools and the standardization of procedures is mandatory to achieve reliable measurements and to ensure their comparability and reproducibility over time, especially in CKD patients in whom body composition can be influenced by various factors related to the disease itself and in particular the hydration state.

Mid-arm circumference (MAC) measurement can be easily applied where other instruments are not available or usable, for example in bedridden patients. MAC is considered a better prognostic indicator of the risk of death associated with malnutrition than the weight-to-height ratio (4). Higher values of MAC and calf circumference (CC) are indicative of greater lean mass and have also been associated with reduced mortality in hemodialysis patients (4,79). Tricipital skin fold (TSF) thickness is the most used single-site skinfold measurement, as it is easily measurable and reference data are available for comparison. There are several equations to derive fat mass from skin fold measurements at different sites, but no equation exists for a single-site skinfold measurement (80-82). Hemodialysis patients with higher BMI and body fat mass seem to have better quality of life and survival (83).

TSF and MAC are used to calculate the middle-arm muscle circumference (MAMC) and the middle-arm muscular area (MAMA) using specific formulas:

\[
MAMC = MAC - (TSF \times \pi)
\]

\[
MAMA = [MAC - (\pi \times TSF)]/4 \pi
\]

Higher MAMC has been found to be predictive of a lower death risk in patients on maintenance dialysis (84). MAMC has been validated versus LBM measured by DXA, a gold standard method for body composition assessment, and a strong correlation has been demonstrated in patients on maintenance hemodialysis, suggesting that MAMC can be considered a valid surrogate of muscle mass (29). MAMC may also reflect adequacy of energy and protein intake, and its reduction can be considered an index of sarcopenia and a sign of malnutrition or wasting (85). The integrated evaluation of MAMC and TSF measurements (MAMC low/high + TSF low/high) may help in the estimation of survival in hemodialysis patients; in this respect, it has been shown that patients with high MAMC combined with either high or low TSF compared to low MAMC and low TSF have the greatest survival probability (29). However, despite its usefulness in clinical practice, because of its intrinsic limitations anthropometry should be complemented by other methods to obtain more accurate results in assessing body composition.

**BIA**

BIA estimates body composition based on the resistance of the body to the passage of an alternating electrical current. Different techniques are available depending on current frequency and/or the site of measurement. On the basis of the current frequencies, two approaches have been developed. In single-frequency bioimpedance analysis (SF-BIA) a low frequency of 50 Hz is delivered, whereas in multifrequency bioimpedance analysis (MF-BIA) the instrument delivers different frequencies that generally vary from 5 to 1,000 kHz or a broadband of frequencies [in this case the technique is called bioimpedance spectroscopy (BIS)]. Based on sites where measurements are performed, BIA is classified as whole body or segmental.

Impedance (Z) represents the force that interferes with the flow of electric current, and is the result of the vectorial sum of the resistance (Rz) and the reactance (Xc). The resistance is the component of the impedance corresponding to the flow of current in fluids, while the reactance is the component of the impedance corresponding to the flow through cell membranes, which behave like capacitors due to their lipid bilayer structure. In the human body, electric
current is conducted by electrolytes present in body fluids, most of which are contained in the fat free mass (FFM). FFM includes bone tissue and body cell mass (BCM), that represents the metabolically active component and includes the SMM. Total body water (TBW), that represents 73% of FFM in normo-hydrated subjects, is the sum of intracellular water (ICW) and extracellular water (ECW).

The most investigated bioelectric index is the phase angle (PA) that derives from a proportion between resistance and reactance according to the following formula:

\[ \text{PA} = \text{Arctang} \left( \frac{Xc}{Rz} \right) \times 180 \times \pi \]  

[3]

The PA is considered the BIA parameter that best predicts survival in CKD and ESKD patients on peritoneal dialysis and maintenance hemodialysis, as well as in patients with HIV, cancer and other diseases (86-93). A PA \( \leq 4.5^\circ \) has been associated with a worse clinical outcome in patients undergoing hemodialysis (94).

As mentioned above, CKD is accompanied by fluid retention and derangements of water distribution between the intra- and the extracellular compartments. In this clinical setting BIA should be considered, as it can detect changes in body fluids, is noninvasive and easy to perform at the bedside (95,96). Using research data obtained in population samples of adequate dimension, nomograms have been developed based only on bioelectric measurements, such as the RXc Graph, that are useful for identifying conditions of adequate or altered hydration state. This nomogram allows a qualitative diagnosis from the resistance and reactance values directly measured by the instrument, using SF-BIA at 50 KHz frequency. Resistance and reactance, normalized for height (H), the length of the conductor (the patient's body), are plotted on the RXc Graph with three elliptical probability regions: 50%, 75%, 95% tolerance ellipses (Figure 2). Rz/H and Xc/H are the two components of the impedance vector. The length of the vector represents the size of the impedance. The movement of the vector along the major axis of the ellipses, gives clinical information on the hydration state and is not influenced by body weight (97).

While SF-BIA is the first and the most used method to assess body compartments, in the last years, body composition monitors based on the application of multiple current frequencies (MF-BIA) have been preferred in clinical research because they estimate separately ICW and ECW (whereas SF-BIA gives information only on ECW), allowing a more precise measurement of body water content. At low frequencies, the current flows through the extracellular fluids, while at high frequencies it can pass through the cells. By changing the current frequency, MF-BIA provides therefore information on the distribution of water outside and inside the cells.

Mathematical models and empirical formulae derived from a reference population are applied to estimate body characteristics. These formulas are derived from a population of healthy subjects belonging to specific ethnic groups; thus, their application in patients with possible alteration of the hydration state and from different ethnic groups could yield biased values. Some authors have shown that MF-BIA is preferable to SF-BIA in the assessment of muscle mass, as it is less affected by over-hydration (98). In hemodialysis patients, estimation of muscle mass and ECW with MF-BIA showed a good correlation with measurements performed with gold standard techniques such as isotopic methods (\( \text{D}_2\text{O} \) and \( \text{K}^{40} \)) and MRI (98,99).

While the hydration state of CKD/ESKD patients may represent an important source of bias in the assessment of muscle mass by BIA, some precautions can be applied to allow greater reliability and reproducibility of this method, both with single or multi-frequency BIA:

- If BIA is used to assess hydration state or to define dry weight, it is recommended that measurements be performed both before and after the dialysis treatment, in order to detect changes in Rz and Xc (the only two parameters really measured by the instruments, as all of the other parameters are estimated by equations);
- If BIA is used to assess muscle mass, it is important to perform the examination after a time interval of 15–120 minutes since the end of the dialysis session,
that is in the dry-weight state, preferably after the second dialysis session of the week (100). In these conditions the mathematical equations provided with the BIA software may give a more reliable estimation of TBW and FFM;

- Regarding peritoneal dialysis patients, some investigators have reported that the presence of dialysis solutions in the peritoneal space has a negligible effect on the assessment of body composition, while others observed an overestimation of body water and cell mass (101,102). Thus, for a more accurate assessment of body composition in peritoneal dialysis patients, it is advisable to perform BIA when the abdomen is empty.

**Imaging methods**

**DXA**

DXA is considered a gold standard method for the assessment of body composition. It is a well-tolerated imaging technique that measures the body relative attenuation of two low-dose X-ray beams with different levels of energy, allowing the identification of different components of body weight, i.e., LBM, fat body mass (FBM) and bone. DXA measurements are based on a three-compartment model that includes total body mineral (from bones), LBM (or lean soft tissue), and fat tissue mass, as opposed to two-compartment models used in SF-BIA; it can also provide both segmental and whole body parameters of body composition.

The recent Sarcopenia, Cachexia and Wasting Disorders position paper recommends DXA as the reference method to assess muscle mass, in particular appendicular skeletal mass, and for the diagnosis of sarcopenia (80). Despite its current and frequent use in clinical practice for the assessment of osteoporosis and sarcopenia in the elderly (5), its accuracy may be affected by hydration state. In fact, DXA assumes that LBM has a constant hydration of 73%, and fluctuations of the hydration state of patients with CKD/ESKD may overestimate or underestimate this parameter (103). Nonetheless, in CKD patients DXA measurements have been associated with physical functioning: measures of adiposity were in fact associated with lower functional ability, whereas higher muscle mass was associated with better physical function and quality of life in dialysis patients (27).

Notwithstanding it not being time-consuming and the minimal patient radiation exposure, evaluation by DXA is not feasible at the bedside, nor does it allow an accurate direct quantification of absolute compartmental tissue volumes. In addition, the cost of the machine and the need for trained radiology operators represent other limitations hampering its routine use for the assessment and monitoring of body composition.

**CT**

CT scan is a noninvasive test generally used for diagnostic and treatment purposes. It can be applied also for the assessment of body composition assessment as it provides high quality images that can differentiate and measure fat and lean body tissue. Moreover, it can distinguish between intra- and extra-abdominal fat, and can also detect the amount of infiltrated fat tissue in SMM, providing information regarding muscle quality. The procedure requires an X-ray source that rotates around the patient emitting a beam of photons. The known differences in attenuations of X-rays between lean soft tissue and adipose tissue are used to separate them, as well as to determine the infiltration of fat in a particular lean soft tissue. Being a three-dimensional technique, CT can be used to perform direct volumetric measurements of organs. However, because CT uses ionizing radiation, CT-based body composition analysis is usually limited to two-dimensional images of one or a limited number of axial slices, allowing for the assessment of muscle area as a proxy to volume and minimizing the ionizing radiation dose. However, this approach limits the precision of the measurement, since the location of the slices may vary between scans.

In the renal population, CT has been applied to assess quadriceps muscle cross-sectional area (CSA) and volume (104,105). In addition, CT has also been used to assess muscle density, which provides additional information regarding muscle quality or the degree of intramuscular fat infiltration (106). Recently, abdominal CT scan has been used in oncology and ICU to assess and monitor changes in muscle mass at the level of the third lumbar vertebra (L3), and may be a promising tool for patients undergoing abdominal CT for diagnostic reasons (107-109). In fact, the axial slice at the level of the third lumbar vertebra showed the highest correlation with total body skeletal muscle volume as assessed by MRI in healthy subjects (110). More recently, in cancer patients, muscle volume assessed by a CT scan at the third lumbar vertebra was highly correlated with FFM and appendicular SMM as assessed by DXA (111). In the CKD population, it has been used to validate surrogate...
methods for the assessment of muscle mass, such as BIA, anthropometry, subjective global assessment (SGA) and two predictive equations in non-dialysis CKD patients (112).

The European Consensus Statement for the diagnosis of sarcopenia (44,108), recommends the use of CT as the gold-standard method for the assessment of muscle loss. In addition, because as it is not influenced by hydration status, it can be used in the CKD/ESKD population with better precision than DXA. However, given the radiation exposure and its high costs, the use of CT for body composition analysis is usually restricted to the research setting.

**MRI**

MRI is a noninvasive imaging technique that uses different properties of the nuclei of hydrogen present in water and fat to produce images of soft tissue in the body. By using quantitative fat water imaging, (or Dixon imaging), precise measurements of adipose and lean tissue, as well as the fat infiltration in organs, can be performed (113). It can give information about body structure and function for diagnostic and treatment purposes. Particularly, in the Dixon imaging, the different magnetic resonance frequencies of protons in fat and water are used to separate water and fat images, facilitating the differentiation of lean and adipose tissue compartments (114). Using this technique, a total body scan with sufficient spatial resolution can take only 6–8 minutes (115). As opposed to CT and DXA, MRI does not use ionizing radiation, which enables its use in a broad population without theoretically increasing the risk of cancer.

Recently published papers have suggested methods that automatically or semi-automatically identify skeletal muscle volumes using MRI (116-120). The fat referenced Dixon imaging protocol has been validated for its accuracy and precision in assessing muscle fat infiltration (121), while Thomas et al. reported test-retest reproducibility for whole body and compartmental muscle volumes with narrow limits of agreement (119).

In the ESKD population, MRI has been used to assess whether frailty and functional tests were associated with body composition (122). In particular, LBM was assessed by DXA, while MRI was used to assess the CSA of the thigh, and BIA was used to measure the PA. Low CSA of the quadriceps muscle and lower PA were associated with worse performance and functional status, while LBM estimated by DXA was not associated with performance and functional status (122). In a more recent study, the psoas muscle area measured at the level of L4-L5, i.e., the same level used for the measurement of abdominal fat content, provided a useful measure of whole-body muscle mass, showing a strong correlation with LBM by DXA (r=0.74; P<0.001), and with the quadriceps muscle CSA (r=0.83; P<0.001) in patients on hemodialysis (123). In addition, this study also validated the MRI-based assessment of the psoas muscle and quadriceps CSA as accurate predictors of sarcopenia (123).

MRI is considered more accurate than CT scans for the assessment of muscle mass because of the increased contrast between tissues with different molecular properties (124). However, just like CT, MRI is expensive and requires specialized personnel. In addition, cut-off values of muscularity are yet to be defined because sex, age, ethnicity and total body fat content are possible factors that influence muscle mass (125). Thus, despite its widespread availability in hospitals, MRI is currently only employed for diagnostic purposes, and its use for the assessment of nutritional status is limited to the research setting.

**Muscle US**

Recently, the application of US scan technique for the assessment of SMM in renal patients has aroused considerable interest. Its major advantages, compared to other modalities, are represented by low cost, portability, lack of radiation exposure and the possibility to be applied by non-specialized staff (6,7,126). Portability is of particular interest in the CKD research setting and clinical practice, since patients can be evaluated during the hemodialysis session or outpatient visits. As no radiation is required, muscle US can be used in every kind of patient, including children and pregnant women. Another important advantage of US is that it allows real-time visualization of the target structure and, through echogenicity, it can provide information about the presence of inflammation, fibrosis, and adipose infiltration (127).

Reliability and validity of US have been clearly documented in the renal setting for the assessment of quadriceps muscle thickness and CSA (6-8,128). Specifically, the reliability of muscle US has been tested in critically ill patients with AKI, showing excellent intraclass correlation coefficients (ICC) for inter- and intra-operator comparisons (6). In the same clinical setting, US assessment of quadriceps muscle has also been validated against CT (8). Differential and proportional bias were small and not statistically significant, while the precision analysis showed slightly lower, though not clinically important, precision for muscle US in comparison to CT scan (8). US was also performed for the assessment of muscle CSA and was validated in CKD patients not on
dialysis (128). In another reliability study, quadriceps muscle thickness values measured by US before and after RRT were not different, suggesting that US is poorly influenced by fluid overload and by rapid and relevant fluid shifts, such as those typical of patients on dialysis (6,7). Despite such encouraging results, more work is needed before assuming muscle US as a reference method for the diagnosis of muscle wasting. Moreover, further studies aimed at defining cut-off values for muscle mass in the general population are needed, in order to allow the early identification of patients with low muscle mass.

Conclusions

Muscle wasting is highly prevalent among CKD/ESKD patients, and is associated with physical disability, worse quality of life and increased morbidity. In this regard, the assessment of nutritional status, and particularly of body composition, are mandatory for an adequate management of renal patients. The choice of the method will depend on its availability and ease of application in clinical practice, local experience, staff resources and good repeatability over time. Anthropometric measures can represent a valid surrogate of body composition parameters if standardized procedures are applied. BIA, another non-invasive and cheap method, despite its being easily performed at the bedside, also requires standard procedures to achieve reliable measurements and ensure their comparability and reproducibility over time, especially in CKD patients. Finally, muscle US, while representing a promising imaging technique as it is simple, easily applicable, valid, accurate and reliable, still needs that more data be accumulated before it can be recommended as a routine tool in clinical practice.

Acknowledgments

Funding: None.

Footnote

Provenance and Peer Review: With the arrangement by the Guest Editors and the editorial office without any funding or sponsorship. GG served as the unpaid Guest Editor of the special issue and serves as an unpaid editorial board member of Quantitative Imaging in Medicine and Surgery. AB served as the unpaid Guest Editor of the special issue. The authors have no other conflicts of interest to declare.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: https://creativecommons.org/licenses/by-nc-nd/4.0/.

References


74. Dwyer JT, Larive B, Leung J, Rocco MV, Greene T,


Quantitative Imaging in Medicine and Surgery, Vol 10, No 8 August 2020


121. Peterson P, Romu T, Brorson H, Dahlqvist Leinhard O, Månsson S. Fat Quantification in Skeletal Muscle Using Multigradient-Echo Imaging: Comparison of Fat and Water...


© Quantitative Imaging in Medicine and Surgery. All rights reserved.  
http://dx.doi.org/10.21037/qims.2020.03.05